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## Stimulation of grassland nitrogen cycling under carbon dioxide enrichment

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**Abstract** Nitrogen (N) limits plant growth in many terrestrial ecosystems, potentially constraining terrestrial ecosystem response to elevated CO<sub>2</sub>. In this study, elevated CO<sub>2</sub> stimulated gross N mineralization and plant N uptake in two annual grasslands. In contrast to other studies that have invoked increased C input to soil as the mechanism altering soil N cycling in response to elevated CO<sub>2</sub>, increased soil moisture, due to decreased plant transpiration in elevated CO<sub>2</sub>, best explains the changes we observed. This study suggests that atmospheric CO<sub>2</sub> concentration may influence ecosystem biogeochemistry through plant control of soil moisture.

**Key words** N mineralization · Elevated CO<sub>2</sub> · Annual grasslands · Soil moisture

### Introduction

Nitrogen limits plant growth in many terrestrial ecosystems (Vitousek and Howarth 1991). Elevated CO<sub>2</sub> could increase or decrease N availability to plants. The conclusion from laboratory and microcosm studies is that increased soil C availability causes these changes. Increased labile C inputs to soil resulting from higher root exudation or turnover under elevated CO<sub>2</sub> can stimulate (Zak et al. 1993) or depress (Díaz et al. 1993) N availability to plants, and stimulate leaching losses of N

(Körner and Arnone 1993). Also, increased C:N ratio in litter produced under elevated CO<sub>2</sub> can slow nutrient release during decomposition (Coûteaux et al. 1991; Field et al. 1992).

Elevated CO<sub>2</sub> can also cause increased soil moisture as a result of decreased plant transpiration (Field et al. 1995). The consequences of increased soil moisture for ecosystem processes, including N cycling (Kuikman et al. 1990; Tietema et al. 1992), have not been considered in elevated CO<sub>2</sub> studies. The work presented here is the first to directly measure changes in ecosystem N cycling in an intact ecosystem under elevated CO<sub>2</sub> and to postulate soil moisture as the driving mechanism.

### Materials and methods

We conducted this research in two grasslands at the Jasper Ridge Biological Preserve of Stanford University, California, United States (37°24'N, 122°14'W; elevation 150 m). The climate is mediterranean, with cool, wet winters and hot, dry summers. Serpentine and sandstone annual grasslands occur adjacent to one another but differ dramatically in productivity and nutrient limitation (Field et al. 1996). On each grassland, open-top chambers maintain either ambient or elevated (ambient+350 ppm) CO<sub>2</sub> atmospheres over ten replicate plots, each covering 0.3-m<sup>2</sup> ground area (Jackson et al. 1994; Field et al. 1996). Here we present results from the first growing season (1992) after establishing the CO<sub>2</sub> treatments.

In April 1992 (when plant biomass is near its seasonal maximum), we measured the gross rates of microbial production and consumption of NH<sub>4</sub><sup>+</sup> and plant NH<sub>4</sub><sup>+</sup> uptake using the <sup>15</sup>N pool dilution technique (Davidson et al. 1991). We added <sup>15</sup>NH<sub>4</sub><sup>+</sup> to each plot by evenly distributing 3 mg <sup>15</sup>N [as 1.5 mmol l<sup>-1</sup> aqueous (<sup>15</sup>NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 99 atom% <sup>15</sup>N] over the soil surface of a 200-cm<sup>2</sup> area in each plot, and then simulating a 5-mm rainfall event to wash the <sup>15</sup>NH<sub>4</sub><sup>+</sup> into the soil. Then, 24 h later, we clipped plant shoots in an 80-cm<sup>2</sup> area centered within the labeled 200-cm<sup>2</sup> area. We removed one 1.9-cm diameter soil core from each plot – 15 cm deep in the sandstone soil and 10 cm deep in the more shallow serpentine. We removed roots from each soil core by hand. We extracted soil solution N from a subsample from each soil core using 0.5 M K<sub>2</sub>SO<sub>4</sub> and immediately froze the extracts. We determined NH<sub>4</sub><sup>+</sup> and NO<sub>3</sub><sup>-</sup> concentrations in the extractions colorimetrically (Lachat 1990). We determined extractable N by Kjeldahl diges-

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tion and  $\text{NH}_4^+$  in the digestion by colorimetry (Lachat 1990). We concentrated the  $\text{NH}_4^+$  from the digested sample using a diffusion technique (Brooks et al. 1989) and then determined  $^{15}\text{N}$  content by combustion isotope-ratio mass spectroscopy.

We calculated gross mineralization using the equations of Kirkham and Bartholomew (1954), with two additional assumptions: (1) that the  $^{15}\text{N}$  recovered from Kjeldahl digestion and diffusion of the non-fumigated extraction was  $^{15}\text{NH}_4^+$ , and (2) that the pool size of soil solution  $\text{NH}_4^+$  did not change. Production of  $^{15}\text{NO}_3^-$  through nitrification during the 24-h labeling period should not violate the first assumption because Kjeldahl digestion typically does not reduce  $\text{NO}_3^-$  (Dalal et al. 1984), and the diffusion procedure collects only  $\text{NH}_4^+$  (Brooks et al. 1989). Also, in similar experiments on the same soils, we have found no differences in the direction or magnitude of change in  $\text{NH}_4^+$  pool size between  $\text{CO}_2$  treatments (B.A. Hungate unpublished work), validating the second assumption. We calculated  $^{15}\text{N}$  enrichments at 0 h as the amount of  $^{15}\text{N}$  added divided by the total  $\text{NH}_4^+$  pool size. We measured  $^{15}\text{N}$  enrichment and N concentration in plants by direct combustion and mass spectroscopy, and in microbes by chloroform-fumigation extraction followed by direct combustion and mass spectroscopy (Davidson et al. 1991). We calculated gross  $\text{NH}_4^+$  uptake rates for plants and microbes using the measured enrichment and an exponential model of  $^{15}\text{N}$  decline in the  $\text{NH}_4^+$  pool during the 24-h period (Davidson et al. 1991). We express gross mineralization and immobilization on a ground area basis, using measured bulk densities of  $0.97 \text{ g soil cm}^{-3}$  for the serpentine and  $1.18 \text{ g soil cm}^{-3}$  for the sandstone (Hungate et al., in press). For each plot, we also calculated the proportion of mineralized  $\text{NH}_4^+$  that plants took up and the proportion that microbes immobilized. In the short-term (24-h) assay, these proportions can be larger than 1 (see e.g., Table 1) due to stimulation of  $^{15}\text{NH}_4^+$  uptake caused by the  $^{15}\text{NH}_4^+$  addition (Davidson et al. 1991).

We calculated total  $^{15}\text{N}$  recovery as the sum of  $^{15}\text{N}$  recovered in soil and plant components at the end of the 24-h period. Across all treatments, we recovered  $88 \pm 4\%$  of the total  $^{15}\text{NH}_4^+$  applied. Two-way analysis of variance (ANOVA) showed that this percentage did not differ between  $\text{CO}_2$  treatments ( $P=0.24$ ) nor between the serpentine and sandstone grasslands ( $P=0.94$ ), and the interaction was not significant ( $P=0.77$ ). Nitrification and subsequent denitrification or leaching of  $^{15}\text{NO}_3^-$  are the likely pathways through which losses of  $^{15}\text{N}$  occurred, reducing  $^{15}\text{N}$  recovery. In these soils, the pH is 5.5–6.5 (Luo et al. in press), and soil  $\text{NH}_4^+$  concentrations in this study were relatively low,  $<2 \mu\text{g N g}^{-1}$  soil (data not shown). Thus, we do not expect  $\text{NH}_3$  volatilization to be an important fate of ammonium.

We measured soil moisture gravimetrically and microbial biomass C using the chloroform-fumigation technique (Voroney and Paul 1984) from the same soil cores used for the  $^{15}\text{N}$  pool dilution. We removed three 5- to 15-g subsamples from each core. In one, we determined gravimetric soil moisture as weight loss after drying for 48 h at  $70^\circ \text{C}$ . We fumigated one of the two remaining subsamples with chloroform vapors for 24 h in a glass desiccator. We incubated each fumigated and non-fumigated subsample in 1-l mason jars for 10 days in the dark. At the end of the incubation, we analyzed the headspace in each jar for  $\text{CO}_2$  concentration by gas chromatography (Shimadzu), then determined microbial biomass C as the difference in  $\text{CO}_2$  production between fumigated and non-fumigated subsamples, divided by 0.41 to correct for C assimilation during the 10-day incubation (Anderson and Domsch 1978).

We determined relative water use efficiency for the four most common species on each grassland using the  $\delta^{13}\text{C}$  technique, using the approach and equations described in Jackson et al. (1994). We measured  $\delta^{13}\text{C}$  by isotope-ratio mass spectroscopy (Europa Scientific, UK) of homogenized shoot material, and calculated carbon isotope discrimination ( $\Delta$ ):

$$\Delta = (\delta_{\text{air}} - \delta_{\text{leaf}}) / (1 + \delta_{\text{air}}),$$

where  $\delta_{\text{air}} = -8\text{‰}$  in ambient  $\text{CO}_2$ . In elevated  $\text{CO}_2$ , the seasonal average  $\delta_{\text{air}}$  was  $-18.4\text{‰}$  (monitored in March, April, and May, and adjusted for plant growth in ambient air before chamber placement on 6 January). Then, we determined  $c_i/c_a$  (the ratio of intercellular

to external  $\text{CO}_2$  concentration) using the equations of Farquhar et al. (1989), and calculated the ratio of water-use efficiencies for two leaves experiencing the same leaf-to-air vapor concentration gradient:

$$(A_1/E_1)/(A_2/E_2) = (c_{a1} - c_{i1}) / (c_{a2} - c_{i2}),$$

where  $A/E$  is the ratio of photosynthesis to transpiration (for leaf 1 or leaf 2). Thus, relative WUE for ambient  $\text{CO}_2$  is defined as 1.

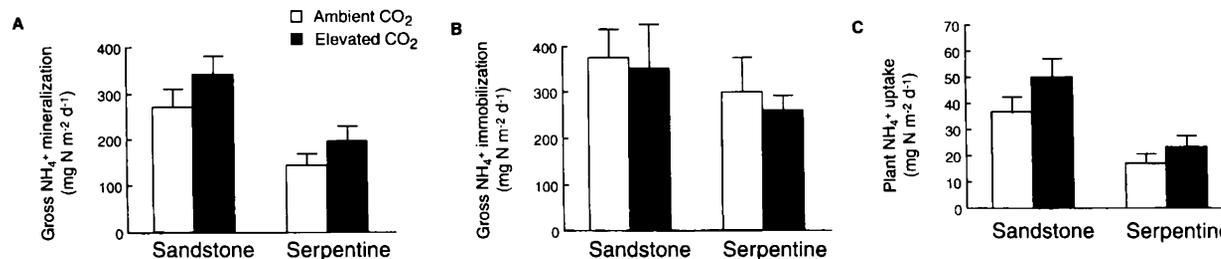
Plant production occurs in two phases in these ecosystems (Chiariello 1989). Early season annuals germinate in October and flower and senesce from March to May. Late season annuals also germinate in October, but flower late in the summer and senesce as late as November. In order to characterize total N uptake by the vegetation and its intraannual pattern, we measured plant N pools for early season (April) and estimated plant N pools for late season production. We calculated total plant N uptake for the early season by summing the product of plant mass (shoots and roots) and N concentration in April. In the serpentine, we estimated late-season annual plant biomass in June using an indirect approach based on plant height:biomass relationships (data from Field et al. 1996). We did not measure N concentration in June, but we have found no evidence for  $\text{CO}_2$  effects on N concentrations in the late season annuals during vegetative growth (April 1992), nor after seed set (January 1994 and 1995; B.A. Hungate, unpublished work). Thus, we calculated N uptake by late-season annuals as biomass in June times N concentration in April; we calculated total N uptake as early plus late-season N uptake.

Standing litter in January includes both early and late season production from the previous growing season. Because we had no separate measure of late season N uptake in the sandstone, we used the total N pool in standing litter in January 1993 as an index of total annual plant N uptake for 1992. We measured total standing dead plant mass in January by clipping plant shoots in an  $80\text{-cm}^2$  in each plot and weighing the dried material; we also measured N concentration by combustion gas-chromatography. We estimated total N uptake as the mass of standing litter times N concentration. We estimated late season N uptake as the difference between total N pool in standing litter in January 1993 and plant N pools in April 1992 (early season). Because some N is lost with seed release and litterfall over the summer, this underestimates both total N uptake and late season N uptake (see e.g., values for the ambient  $\text{CO}_2$  treatment, Table 2). However, this approach provides an index of the effect of elevated  $\text{CO}_2$  on total plant N uptake during the 1992 growing season.

When we had comparable measurements in the two ecosystems, we tested for the effects of elevated  $\text{CO}_2$  using two-way ANOVA with  $\text{CO}_2$  and ecosystems as the main effects. To test for  $\text{CO}_2$  effects within each ecosystem for late season plant N uptake, we used single factor ANOVAs. We used analysis of covariance (ANCOVA) to investigate the potential mechanisms of the effects of elevated  $\text{CO}_2$  on gross mineralization, using soil moisture and microbial biomass C as covariants. For this analysis, we used data expressed on a per gram soil basis for all three variables.

## Results and discussion

Elevated  $\text{CO}_2$  caused an increase in the gross rate of  $\text{NH}_4^+$  mineralization on both serpentine and sandstone grasslands (two-way ANOVA,  $P=0.088$ , Fig. 1A) when plants were approaching their maximum biomass. Elevated  $\text{CO}_2$  did not alter gross microbial  $\text{NH}_4^+$  immobilization (two-way ANOVA,  $P=0.866$ , Fig. 1B). The proportion of mineralized  $\text{NH}_4^+$  that was immobilized (two-way ANOVA,  $P=0.075$ , Table 1) decreased in elevated  $\text{CO}_2$ , indicating greater  $\text{NH}_4^+$  availability. In no case was the  $\text{CO}_2$  by ecosystem interaction significant (two-way ANOVAs,  $P>0.57$ , Fig. 1).



**Fig. 1** A Gross NH<sub>4</sub><sup>+</sup> mineralization, B gross NH<sub>4</sub><sup>+</sup> immobilization, and C plant NH<sub>4</sub><sup>+</sup> uptake for 2 April 1992 in serpentine and sandstone grasslands in ambient and elevated CO<sub>2</sub> treatments. Values presented are means±standard errors (n=10)

Plant NH<sub>4</sub><sup>+</sup> uptake increased in elevated CO<sub>2</sub> (two-way ANOVA, *P*=0.081, Fig. 1C). Though this could occur as a result of a direct CO<sub>2</sub> stimulation of plant NH<sub>4</sub><sup>+</sup> uptake, elevated CO<sub>2</sub> did not alter the proportion of mineralized N that plants took up (two-way ANOVA, *P*=0.69, Table 1). This suggests that the increase in plant NH<sub>4</sub><sup>+</sup> uptake occurred in response to greater NH<sub>4</sub><sup>+</sup> availability caused by increased NH<sub>4</sub><sup>+</sup> mineralization. Increased plant NH<sub>4</sub><sup>+</sup> uptake accounts for only a part of the extra NH<sub>4</sub><sup>+</sup> mineralized in elevated CO<sub>2</sub>. While NH<sub>4</sub><sup>+</sup> mineralization increased by 53–60 mg N m<sup>-2</sup> day<sup>-1</sup> in elevated CO<sub>2</sub> (Fig. 1, Table 1), plant uptake only increased by 5–13 mg N m<sup>-2</sup> day<sup>-1</sup> (Fig. 1, Table 1). Thus, we cannot account for 75–92% of the extra NH<sub>4</sub><sup>+</sup> mineralized in elevated CO<sub>2</sub>. Nitrification and subsequent N losses through nitrate leaching and denitrification are plausible fates for this NH<sub>4</sub><sup>+</sup> (Robertson 1989).

Elevated CO<sub>2</sub> had no effect on total plant N uptake during the early part of the growing season (two-way ANOVA, *P*=0.96, Table 2) but substantially increased N uptake by the late season vegetation in the serpentine (one-way ANOVA, *P*=0.04, Table 2) and N pools in litter in January, our index of total N uptake in the sandstone (one-way ANOVA, *P*=0.01, Table 2). Hence, elevated CO<sub>2</sub> increased plant N acquisition late in the growing season.

We observed no dilution of N in plant tissue in elevated CO<sub>2</sub> in late-season annuals during vegetative growth (two-way ANOVA, *P*=0.98, Table 2), and found no effect of elevated CO<sub>2</sub> on N concentrations in litter at the end of the growing season (two-way ANOVA, *P*=0.99, Table 2). Thus, the increase in N uptake shown here is proportional to the CO<sub>2</sub> stimulation of late-season plant production (Field et al. 1996): these plants are able to obtain sufficient N to meet increased growth in elevated CO<sub>2</sub> and do not show signs of increased N stress. A sustained increase in the availability of NH<sub>4</sub><sup>+</sup> – the dominant source of N for these annual plants (Jackson et al. 1989; Jackson and Reynolds 1996) – is the likely cause of the increased annual N uptake by plants with no dilution of N in plant tissue.

**Table 1** The proportion of gross NH<sub>4</sub><sup>+</sup> mineralization that plants take up and that microbes immobilize, and absolute changes in plant NH<sub>4</sub><sup>+</sup> uptake and gross NH<sub>4</sub><sup>+</sup> mineralization in response to elevated CO<sub>2</sub>. Elevated CO<sub>2</sub> did not alter the proportion of mineralized NH<sub>4</sub><sup>+</sup> that plants take up, but reduced the proportion of min-

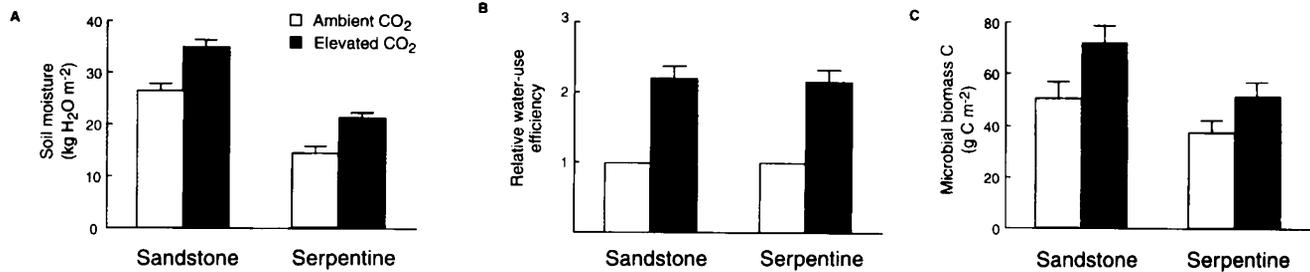
eralized NH<sub>4</sub><sup>+</sup> that is subsequently immobilized. Values for proportions are means±SEM (n=10). Values for absolute changes in response to elevated CO<sub>2</sub> are the differences in means (n=10) between elevated and ambient CO<sub>2</sub> treatments

Ecosystem	CO <sub>2</sub>	Proportion of gross mineralization as		Increase in elevated CO <sub>2</sub> (in mg N m <sup>-2</sup> day <sup>-1</sup> )	
		Plant uptake	Immobilization	Plant uptake	Mineralization
Serpentine	Ambient	0.17±0.05	2.93±0.83	4.6	59.3
	Elevated	0.11±0.02	1.48±0.26		
Sandstone	Ambient	0.15±0.02	1.53±0.30	12.8	52.5
	Elevated	0.17±0.03	1.19±0.33		

**Table 2** N concentration (g N per 100 g plant) in late-season annuals during vegetative growth (April) and in litter at the end of the 1992 growing season (January 1993), and early season, late season, and total aboveground plant N uptake for 1992 (g N m<sup>-2</sup>). Elevated

CO<sub>2</sub> caused a 50% stimulation of aboveground N uptake in serpentine and sandstone grasslands. Increased N uptake occurred during the late part of the growing season, the time when the effects of elevated CO<sub>2</sub> on increased soil moisture are most pronounced

Ecosystem	CO <sub>2</sub>	N concentration (%)		Plant N uptake for 1992 (g N m <sup>-2</sup> )		
		Late annuals in April	Litter in January	Early N uptake	Late N uptake	Annual N uptake
Serpentine	Ambient	1.33±0.08	0.97±0.03	1.38±0.09	0.47±0.15	1.85±0.17
	Elevated	1.31±0.06	0.97±0.05	1.35±0.08	1.37±0.37	2.73±0.42
Sandstone	Ambient	1.37±0.13	0.74±0.02	2.55±0.49	0	2.10±0.18
	Elevated	1.39±0.06	0.74±0.04	2.33±0.45	0.85	3.18±0.34



**Fig. 2** A Soil moisture, B relative water use efficiency, and C microbial biomass carbon for 2 April 1992. Values presented are means  $\pm$  standard errors (A and C,  $n=10$ ; B  $n=4$ )

We investigated the possible mechanism for increased  $\text{NH}_4^+$  availability in April 1992, the time of peak ecosystem biomass. Soil moisture was 40% higher in the elevated  $\text{CO}_2$  treatment in both sandstone and serpentine grasslands (two-way ANOVA,  $P < 0.001$ , Fig. 2A). This reflects decreased stomatal conductance and transpiration (Jackson et al. 1994), and a doubling of whole plant water-use efficiency in the dominant plants on both grasslands (Fig. 2B). Decreased transpiration per unit of leaf area could be compensated by increased leaf area or increased evaporation from the soil surface, but these compensations, if they occurred, were too small to eliminate the effect of elevated  $\text{CO}_2$  on soil moisture. Elevated  $\text{CO}_2$  also increased microbial biomass C by 27% on the serpentine and by 48% on the sandstone (two-way ANOVA,  $P = 0.011$ , Fig. 2C). Thus, there was evidence that  $\text{CO}_2$  altered both soil moisture and soil C availability.

Analysis of covariance provides correlative support for the hypothesis that increased soil moisture caused the increase in gross  $\text{NH}_4^+$  mineralization we observed. The main effect of  $\text{CO}_2$  on gross  $\text{NH}_4^+$  mineralization was not significant (two-way ANCOVA,  $P = 0.607$ ) when soil moisture and microbial biomass C were included as covariants, indicating that they accounted for the  $\text{CO}_2$  effect. However, whereas microbial biomass C was not a significant covariant for gross  $\text{NH}_4^+$  mineralization (two-way ANCOVA,  $P = 0.246$ ), soil moisture was a significant covariant (two-way ANCOVA,  $P = 0.016$ ). The stronger relationship between soil moisture and gross mineralization (Pearson correlation,  $r = 0.435$ ,  $P = 0.005$ ) than between microbial biomass C and gross mineralization (Pearson correlation,  $r = 0.070$ ,  $P = 0.668$ ) suggests that increased soil moisture is more likely to be the cause of the higher gross mineralization in elevated  $\text{CO}_2$ .

These results cannot unequivocally show that increased soil moisture is the mechanism causing increased gross mineralization because we did not independently manipulate soil moisture and  $\text{CO}_2$  concentration. Nevertheless, we suggest that increased soil moisture in elevated  $\text{CO}_2$  is a plausible and simple explanation for the changes we observed, and that our ANCOVA analysis provides correlative support for this hypothesis. Increased soil moisture can enhance bacterial motility and accessibility to substrates (Hamdi 1971), stimulate protozoan grazing and associated N mineralization (Kukman et al. 1991), and increase substrate diffusion (Da-

vidson et al. 1990). Any one or combination of these could explain the results we observed.

The changes in N cycling under elevated  $\text{CO}_2$  were qualitatively similar in serpentine and sandstone grasslands, though these grasslands differ in many characteristics (Field et al. 1996). The similar patterns in contrasting grassland ecosystems suggest that stimulated N cycling resulting from increased soil moisture under elevated  $\text{CO}_2$  may occur in many grassland ecosystems, and possibly in other ecosystems where plant canopies control soil moisture. To date, increased soil moisture or decreased plant water stress under elevated  $\text{CO}_2$  has been observed in grassland (Field et al. 1995) and agricultural forb (Clifford et al. 1993) ecosystems, and it is likely to occur in a broad range of water-limited grassland, shrubland, and forest ecosystems (Field et al. 1995). Repeated monitoring at Jasper Ridge using time-domain reflectometry (Topp et al. 1980) in 1993–1995 indicates consistently increased soil moisture in the sandstone grassland under elevated  $\text{CO}_2$ , both late in the growing season and during midseason dry spells (Field et al. 1995). Some of the measurements indicate smaller  $\text{CO}_2$  effects on soil moisture on serpentine than sandstone (A.L. Fredeen, C.P. Lund, and C.B. Field, unpublished work), perhaps as a result of greater evaporation in the serpentine, with a large amount of bare soil (Schulze et al. 1994).

The long-term effects of elevated  $\text{CO}_2$  on plant production will depend on a number of factors, including mechanisms that operate on a longer time scale than this study addressed. The direct  $\text{CO}_2$  stimulation of photosynthesis is the primary mechanism favoring increased plant production, but nutrient limitation may counteract this response by constraining the  $\text{CO}_2$  stimulation of production from the outset, and by exacerbating N limitation through decreased litter quality and reduced N availability to plants (Mooney et al. 1991). Also, increased turnover of roots and exudation of labile C in elevated  $\text{CO}_2$  can alter N availability (Zak et al. 1993; Díaz et al. 1993; Körner and Arnone 1993), though the importance of this in natural ecosystems is unknown.

In this study, we suggest that elevated  $\text{CO}_2$  stimulated soil N mineralization through increased soil moisture. Though we can not rule out the possibility that increased N mineralization and soil moisture also enhanced N losses (Davidson and Swank 1986; Robertson 1989), we found that greater N mineralization in elevated  $\text{CO}_2$  led to increased plant N uptake. In N-limited ecosystems, this  $\text{CO}_2$ -stimulation of N mineralization through increased soil moisture could release plant growth from N limitation and stimulate plant production.

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## References

- Anderson JPE, Domsch KH (1978) Mineralization of bacteria and fungi in chloroform-fumigated soils. *Soil Biol Biochem* 10:207–213
- Brooks PD, Stark JM, McInteer BB, Preston T (1989) Diffusion method to prepare soil extracts for automated nitrogen-15 analysis. *Soil Sci Soc Am J* 53:1707–1711
- Chiariello NR (1989) Phenology of California grassland. In: Huehneke LF, Mooney HA (eds) *Grassland structure and function: California annual grassland*. Kluwer, Dordrecht, pp 47–58
- Clifford SC, Stronach IM, Mohamed AD, Azam-Ali SN, Crout NMJ (1993) The effects of elevated atmospheric carbon dioxide and water stress on light interception, dry matter production and yield in stands of groundnut (*Arachis hypogaea* L.). *J Exp Bot* 44:1763–1770
- Coûteaux MM, Mousseau M, Celerier ML, Bottner P (1991) Increased atmospheric CO<sub>2</sub> and litter quality: decomposition of sweet chestnut leaf litter with animal food webs of different complexities. *Oikos* 61:54–64
- Dalal RC, Sahrawat KL, Myers RJK (1984) Inclusion of nitrate and nitrite in the Kjeldahl nitrogen determination of soils and plant material using sodium thiosulfate. *Comm Soil Sci Plant Anal* 15:1453–1461
- Davidson EA, Swank WT (1986) Environmental parameters regulating gaseous nitrogen losses from two forested ecosystems via nitrification and denitrification. *Appl Environ Microbiol* 52:1287–1292
- Davidson EA, Stark JM, Firestone MK (1990) Microbial production and consumption of nitrate in an annual grassland. *Ecology* 71:1968–1975
- Davidson EA, Hart SC, Shanks CA, Firestone MK (1991) Measuring gross nitrogen mineralization, immobilization, and nitrification by <sup>15</sup>N isotopic pool dilution in intact soil cores. *J Soil Sci* 42:335–349
- Díaz SA, Grime JP, Harris J, McPherson E (1993) Evidence of a feedback mechanism limiting plant response to elevated carbon dioxide. *Nature* 364:616–617
- Farquhar GD, Ehleringer JR, Hubick (1989) Carbon isotope discrimination and photosynthesis. *Annu Rev Plant Physiol* 40:503–537
- Field C, Chapin FS III, Matson PA, Mooney HA (1992) Responses of terrestrial ecosystems to the changing atmosphere: a resource-based approach. *Annu Rev Ecol Syst* 23:201–235
- Field CB, Jackson RB, Mooney HA (1995) Stomatal responses to increased CO<sub>2</sub>: Implications from the plant to the global scale. *Plant Cell Environ* 18:1214–1225
- Field CB, Chapin FS III, Chiariello NR, Holland EA, Mooney HA (1996) The Jasper Ridge CO<sub>2</sub> experiment: design and motivation. In: Koch GW, Mooney HA (eds) *Carbon dioxide and terrestrial ecosystems*. Academic Press, San Diego, pp 121–145
- Fredeen AL, Randerson JT, Holbrook NM, Field CB (submitted) Elevated atmospheric CO<sub>2</sub> increases late-season water availability in a water-limited grassland ecosystem. *Plant Cell Environ*
- Hamdi YA (1971) Soil-water tension and the movement of rhizobia. *Soil Biol Biochem* 3:121–126
- Hungate BA, Jackson RB, Field CB, Chapin FS III (in press) Detecting changes in soil carbon in CO<sub>2</sub> enrichment experiments. *Plant Soil*
- Jackson RB, Reynolds HL (1996) Nitrate and ammonium uptake for single- and mixed-species communities grown at elevated CO<sub>2</sub>. *Oecologia* 105:74–80
- Jackson LE, Schimel JP, Firestone MK (1989) Short-term partitioning of ammonium and nitrate between plants and microbes in an annual grassland. *Soil Biol Biochem* 21:409–415
- Jackson RB, Sala OE, Field CB, Mooney HA (1994) CO<sub>2</sub> alters water use, carbon gain, and yield for the dominant species in a natural grassland. *Oecologia* 98:257–262
- Kirkham D, Bartholomew WV (1954) Equations for following nutrient transformation in soil, utilizing tracer data. *Soil Sci Soc Am Proc* 18:33–34
- Körner C, Arnone JA III (1993) Responses to elevated carbon dioxide in artificial tropical ecosystems. *Science* 257:1672–1675
- Kuikman PJ, Jansen AG, Veen JA van, Zehnder AJB (1990) Protozoan predation and the turnover of soil organic carbon and nitrogen in the presence of plants. *Biol Fert Soil* 10:22–28
- Kuikman PJ, Jansen AG, Veen JA van (1991) <sup>15</sup>N-nitrogen mineralization for bacteria by protozoan grazing at different soil moisture regimes. *Soil Biol Biochem* 23:193–200
- Lachat (1990) Operation manual for the QuikChem automated ion analyzer. Lachat Instruments, Milwaukee
- Luo Y, Jackson RB, Field CB, Mooney HA (in press) Elevated CO<sub>2</sub> increases belowground respiration in California grasslands. *Oecologia*
- Mooney HA, Drake BG, Luxmoore RJ, Oechel WC, Pitelka LF (1991) Predicting ecosystem responses to elevated CO<sub>2</sub> concentrations. *BioScience* 41:96–104
- Robertson GP (1989) Nitrification and denitrification in humid tropical ecosystems: potential controls on nitrogen retention. In: Proctor J (ed) *Mineral nutrients in tropical forest and Savanna Ecosystems*. Blackwell, Oxford, pp 55–69
- Schulze E-D, Kelliher FM, Körner C, Lloyd J, Leuning R (1994) Relationship among maximum stomatal conductance, ecosystem surface conductance, carbon assimilation rate, and plant nitrogen nutrition. *Annu Rev Ecol Syst* 25:629–600
- Tietema A, Warmerdam B, Lenting E, Riemer L (1992) Abiotic factors regulating nitrogen transformations in the organic layer of acid forest soils: moisture and pH. *Plant Soil* 147:69–78
- Topp GC, Davis JL, Annan AP (1980) Electromagnetic determination of soil water content: measurement in coaxial transmission lines. *Water Resources Res* 16:574–582
- Vitousek PM, Howarth RW (1991) Nitrogen limitation on land and in the sea: how can it occur? *Biogeochemistry* 13:87–115
- Voroney RP, Paul EA (1984) Determination of k<sub>C</sub> and k<sub>N</sub> in situ for calibration of the chloroform fumigation-incubation method. *Soil Biol Biochem* 16:4–14
- Zak DR, Pregitzer KS, Curtis PS, Teeri JA, Fogel R, Randlett DA (1993) Elevated atmospheric CO<sub>2</sub> and feedback between carbon and nitrogen cycles. *Plant Soil* 151:105–117