

Research review

Below-ground process responses to elevated CO₂ and temperature: a discussion of observations, measurement methods, and models

Author for correspondence:

Elise Pendall

Tel: +1 307 766 6293 Fax: +1 307 766 2851 Email: Pendall@uwyo.edu

Received: 14 November 2003 Accepted: 30 January 2004

doi: 10.1111/j.1469-8137.2004.01053.x

Elise Pendall¹, Scott Bridgham², Paul J. Hanson³, Bruce Hungate⁴, David W. Kicklighter⁵, Dale W. Johnson⁶, Beverly E. Law⁷, Yiqi Luo⁸, J. Patrick Megonigal⁹, Maria Olsrud¹⁰, Michael G. Ryan¹¹ and Shiqiang Wan¹² ¹Department of Botany, University of Wyoming, Laramie, WY 82071, USA; ²Center for Ecology and Evolutionary Biology, University of Oregon, Eugene, OR 97403, USA; ³Environmental Sciences

Division, Oak Ridge National Laboratory, Oak Ridge, TN 37831, USA; ⁴Department of Biological Sciences, Box 5640, Northern Arizona University, Flagstaff, AZ 86011, USA; ⁵The Ecosystems Center, Marine Biological Laboratory, Woods Hole, MA 02543, USA; ⁶Department of Natural Resources and Environmental Science, University of Nevada, Reno, Reno, NV 89557, USA; ⁷Department of Forest Science, Oregon State University, Corvallis, OR 97331, USA; ⁸Department of Botany and Microbiology, University of Oklahoma, 770 Van Vleet Oval, Norman, OK 73019, USA; ⁹Smithsonian Environmental Research Center, PO Box 28, 640 Contees Wharf Road, Edgewater, MD 21037, USA; ¹⁰Geobiosphere Science Centre, Lund University, 223 62 Lund, Sweden; ¹¹USDA Forest Service Rocky Mountain Research Station 240 West Prospect Rd., Fort Collins, CO 80526, USA and Department of Forest Rangeland and Watershed Stewardship and Graduate Degree Program in Ecology, Colorado State University, Fort Collins, CO 80523, USA; ¹²Institute of Botany, the Chinese Academy of Sciences, Xiangshan, Beijing 100093, China, and Environmental Sciences Division, Oak Ridge National Laboratory, Oak Ridge, TN 37831, USA

Summary

Key words: carbon sequestration, CO₂ fertilization, mycorrhizas, nutrient cycling, rhizosphere, soil carbon, soil respiration, soil warming.

Rising atmospheric CO_2 and temperatures are probably altering ecosystem carbon cycling, causing both positive and negative feedbacks to climate. Below-ground processes play a key role in the global carbon (C) cycle because they regulate storage of large quantities of C, and are potentially very sensitive to direct and indirect effects of elevated CO_2 and temperature. Soil organic matter pools, roots and associated rhizosphere organisms all have distinct responses to environmental change drivers, although availability of C substrates will regulate all the responses. Elevated CO_2 increases C supply below-ground, whereas warming is likely to increase respiration and decomposition rates, leading to speculation that these effects will moderate one another. However, indirect effects on soil moisture availability and nutrient supply may alter processes in unexpected directions. Detailed, mechanistic understanding and modelling of below-ground flux components, pool sizes and turnover rates is needed to adequately predict long-term, net C storage in ecosystems. In this synthesis, we discuss the current status of below-ground responses to elevated CO_2 and

temperature and potential feedback effects, methodological challenges, and approaches to integrating models and measurements.

© New Phytologist (2004) **162**: 311–322

Introduction

In the coming century, atmospheric CO₂ concentrations are expected to double, and global average temperature may increase by 1.8–5.8°C (IPCC, 2001). The terrestrial C cycle is already probably changing in response to these perturbations (e.g. Raich *et al.*, 2002; Lenton & Huntingford, 2003), but substantial uncertainties remain in the sensitivity of ecosystems to global change forcing factors, particularly regarding the role of feedback among key processes. The capacity of ecosystems to store C depends on net ecosystem production (NEP), which is the balance between net primary production (NPP) and heterotrophic respiration (Rh). Knowledge of the underlying

mechanisms driving changes in NEP is essential to predict terrestrial C cycle responses to rising temperature and CO₂. Current understanding suggests that the primary direct ecosystem response to increased CO₂ concentration is an increase of NPP (i.e. 'CO₂ fertilization'), which is potentially a negative feedback on atmospheric CO₂ concentrations. Rising temperatures could exert their strongest influence over microbial processes such as heterotrophic respiration, which would be a positive feedback on atmospheric CO₂ (Fig. 1a). Simulations suggest that these effects may nearly counteract one another (Kirschbaum, 2000). However, both elevated CO₂ and warming have other direct and indirect effects, which make it unlikely that the primary direct effects will simply

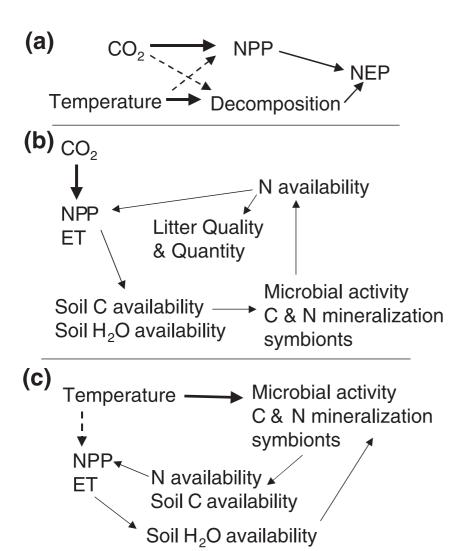


Fig. 1 Direct and indirect effects of elevated CO₂, temperature, and their interactions on C cycling below-ground. (a) Elevated CO2 directly stimulates net primary production (NPP) by enhancing the efficiency of Rubisco, which increases net ecosystem production (NEP). Increased temperature directly stimulates decomposition rates by enhancing enzyme activity and chemical reaction rates, thereby reducing NEP. Elevated CO₂ indirectly affects decomposition rate, either suppressing it via decreased litter quality or enhancing it via the priming effect (see text). Temperature has smaller direct effects on NPP than atmospheric CO2 concentration; interactions and indirect effects make predicting C storage (NEP) challenging. (b) Feedbacks effected by elevated CO2 include water and nutrient cycling. As NPP is stimulated, more C is available in soils, feeding substrate limited microbes. Enhanced microbial activity stimulates C and N mineralization and increases N availability to a limit. Activity of symbionts is likely to be affected by increased C availability below-ground. Litter quantity increases, and as N becomes immobilized in microbes, litter quality may decrease. Elevated CO₂ reduces evapotranspiration (ET) and increases available soil water in some systems, which may enhance NPP and mineralization rates. (c) Increased temperature most strongly stimulates microbial activity, increasing N availability and NPP. In some systems, enhanced ET dries soils, and microbial activity is not stimulated. Temperature may have strong effects on symbiotic interactions.

cancel out; for example, elevated CO₂ also affects microbial activities, and temperature influences NPP (Fig. 1a). This complex web of interactions and feedbacks is slowly being untangled by experiments and modelling to improve our understanding of the combined effects of elevated CO₂ and temperature on the global C cycle.

Direct effects of elevated CO₂ on below-ground C cycling include stimulation of root growth (below-ground net primary production, BNPP) and respiration, increased C inputs from canopy litter fall and root turnover, and changes in litter quality or decomposability (Norby, 1994; Norby et al., 2001, Fig. 1b). Direct effects of increased temperature accelerate losses of CO₂ and CH₄ from the soil by increasing the activity of roots and soil heterotrophs (Hobbie, 1996; Van Cleve et al., 1990; Joslin & Wolfe, 1993; Peterjohn et al., 1994; Lükewille & Wright, 1997). Higher temperatures are also associated with increased NPP, potentially providing more substrate for heterotrophs in the long term if other resources are not limiting (Jenkinson et al., 1991; Raich & Schlesinger, 1992; Lloyd & Taylor, 1994; Kirschbaum, 1995; Trumbore, 1997, Fig. 1c).

Indirect effects of both elevated CO₂ and temperature on the below-ground C cycle are mediated through nutrient and water cycles. Nitrogen (N) mineralization may be stimulated by warming and act as a positive feedback to plant productivity (Strömgren & Linder, 2002). Increased temperature may dry upland soils, which could result in immobilization of essential elements and reduced decomposition; in wetland soils drying may increase aerobic respiration and decomposition by lowering water tables. However, drying may be alleviated if elevated CO₂ reduces transpiration rates (Megonigal & Schlesinger, 1997; Jackson et al., 1998). Plant community shifts will mediate some of these feedbacks: in a montane meadow warming experiment, soil drying induced a shift from productive forbs to less productive shrubs. Warming thus decreased net C uptake, not because decomposition was stimulated, but because NPP was reduced (Saleska et al., 1999). Finally, the net effect of warming and elevated CO₂ on radiative forcing is determined by indirect effects on the production and release of other trace gases such as N₂O and CH₄ (Dacey et al., 1994; Smart et al., 1997; McLain et al., 2002). Separation of temperature, CO₂, moisture and nutrient effects on below-ground processes is a prerequisite for predictive understanding of ecosystem C cycling. The complexity of the interacting processes and their spatial and temporal variability requires greater integration of experimental manipulations and modelling.

The goals of this synthesis are: (i) summarize the current status of the combined effects of experimentally elevated CO₂ and temperature on below-ground processes; (ii) highlight the role of feedbacks mediated through the N and water cycles; (iii) discuss methodological challenges; and (iv) suggest approaches for integrating models and measurements. Below-ground processes include C allocation below-ground via roots; microbial and mycorrhizal processes; SOM pool sizes and turnover rates; and soil, microbial and rhizosphere respiration rates. Although our ability to predict responses of below-ground processes to altered climate is still limited by a lack of longterm, multifactor experiments, understanding of basic mechanisms has greatly improved over the last 10 years. Here we focus on what we believe are the most important recent developments and future directions for below-ground ecosystem

Response of Below-ground C Pools and Processes to Elevated CO₂, Warming and their Interactions

Root production and turnover rates

Fine roots are a key link for plant water and nutrient uptake, soil C input, and soil microbial activity (Fig. 2; Norby, 1994). Turnover of fine roots (< 2.0 mm in diameter) plays a critical role in regulating ecosystem C balance, and accurate estimates of below-ground NPP are required for estimating NEP (Pendall et al., 2004a). It is estimated that as much as 33% of global annual NPP is used for the production of fine roots (Jackson et al., 1997). With their high turnover rate, fine roots will be sensitive to elevated atmospheric CO₂, temperature and their interactions (Hendrick & Pregitzer, 1992; Raich & Schlesinger, 1992; Fitter et al., 1997; Eissenstat et al., 2000; Gill & Jackson, 2000), and may influence sequestration of atmospheric CO₂ on annual to decadal timescales.

Increased photosynthesis under elevated CO₂ can stimulate below-ground C input and fine-root growth (Curtis, 1996; Curtis & Wang, 1998; Pendall et al., 2004b), and root turnover rates and biomass (Berntson & Bazzaz, 1997; Fitter et al., 1999; Allen et al., 2000; Pregitzer et al., 2000; Wan et al., 2004). Higher temperatures are associated with increased fine-root production and mortality (Gill & Jackson, 2000), and therefore turnover rates (Hendrick & Pregitzer, 1993, 1997; Forbes et al., 1997; Fitter et al., 1999; King et al., 1999; Wan et al., 2004). A few experiments have evaluated root responses to the interactive effects of CO₂ and temperature. A 4-year open-top chamber experiment in Tennessee showed that both the main effects of elevated CO₂ and temperature on root turnover (productivity and mortality) of two deciduous tree species were statistically significant and additive; i.e. there were no interactive effects of CO2 and temperature (Wan et al., 2004). The combination of elevated CO₂ and temperature significantly increased fine-root biomass of Loblolly pine seedlings (*Pinus taeda*), but had no effect on Ponderosa pine seedlings (*P. ponderosa*; King et al., 1996). By contrast, Soussana et al. (1996) and Kandeler et al. (1998) found significant reductions in root biomass under elevated CO₂ and temperature. Apparently, elevated CO₂ can sometimes compensate for the anticipated negative effects of increased temperature on root biomass (Bassow et al., 1994; Wan et al., 2004), possibly by reducing evapotranspiration and increasing soil moisture (Nelson et al., 2004), by increasing the

Fig. 2 Below-ground carbon cycle processes and components affected by elevated CO_2 , warming, and their interactions. Although soil organic matter is a complex mixture of materials spanning a continuum of decomposability, we conceptualize three main pools, Active, Slow and Passive. The Active pool receives inputs from the rhizosphere and above-ground litter, and turns over on time scales of up to a few years. The Slow pool receives most inputs from the active pool, with remineralization of Passive C contributing small amounts, and turns over on decadal to century time scales. The Passive C pool consists of physically or chemically protected organo-mineral complexes, with turnover times of millennia. CO_2 efflux is derived from decomposition of the various C pools, including roots and litter, and varies with soil temperature, moisture, and plant phenology. Erosion of particulate organic matter and leaching of dissolved organic carbon are important fluxes in many systems. Decomposition of C pools in all soils produces both CH_4 and CO_2 , with the ratio of the two regulated by O_2 and saturation. The CH_4/CO_2 ratio ranges from zero in dry upland soils where production is restricted to microsites, to perhaps 1/3 in permanently wet, hydric soils; in some systems the ratio varies seasonally or spatially. Methane production is more dependent on labile carbon pools than is CO_2 production, but there is generally less known about the contribution of various C pools to CH_4 production than CO_2 production.

erosion

temperature optimum for photosynthesis (Long, 1991) or by providing carbohydrate supplies to support sustained root growth. Root responses will be partly determined by the growth stage of the experimental plants; larger positive responses may be detected in young systems where the soil volume is not fully occupied.

Root turnover and respiration rates are positively correlated with fine root N concentration (Ryan et al., 1996; Pregitzer et al., 1998, 2002; Eissenstat et al., 2000), which is expected to decrease by 10–25% under elevated CO₂ (Curtis et al., 1990; Berntson & Bazzaz, 1997; King et al., 1997; Cotrufo et al., 1998; Rogers et al., 1999; Pregitzer et al., 2000; Wan et al., 2004). By contrast, elevated temperatures are reported to increase root N concentration (King et al., 1997; Kandeler et al., 1998; Wan et al., 2004), presumably because mineralization and diffusion of N are stimulated at high temperatures (BassiriRad et al., 1993; BassiriRad, 2000). Higher root N concentration could lead to greater fine-root mortality (Pregitzer et al., 1998, 2002), which has the potential to alter rates

of microbial immobilization and modify soil N cycling (Zak et al., 2000a).

Very few studies have reported responses of root N concentration to CO_2 and temperature interactions (King *et al.*, 1997; Loiseau & Soussana, 1999; Wan *et al.*, 2004). Wan *et al.* (2004) found that the interactions of elevated CO_2 and temperature significantly increased root N concentration, but King *et al.* (1997) found no interactive effects. Studies that measure separate rhizosphere respiration responses are required to assess how N concentration and root maintenance cost respond to the interactions of elevated CO_2 and temperature.

Microbial biomass and community structure

Most important biogeochemical processes in soil are microbially mediated, so it is imperative to understand how soil microbial biomass will respond to warming and enhanced CO₂. Although increased labile C input to soil under elevated CO₂ is expected to increase microbial C and N, increases, decreases, and neutral

responses have been found (Zak et al., 2000b). Clear relationships with NPP are lacking, despite the strong relationship between plant productivity and microbial biomass when comparing different ecosystem types (Zak et al., 1993). Negative and neutral microbial biomass responses may be explained by increased turnover rates if soil water content is high (Hungate et al., 1997; Arnone & Bohlen, 1998), if grazing by soil organisms is stimulated (Jones et al., 1998; Ronn et al., 2002) or N availability is low (Diaz et al., 1993). Microbial biomass generally responds positively to increased temperature, whereas responses to elevated CO₂ are highly idiosyncratic. Possibly, microbial response to the interaction of CO₂ and temperature would be dominated by the more ubiquitous temperature effect, or as shown below for mycorrhizas, the effects may offset one another. The community structure of free-living and symbiotic microbes is likely to mediate belowground C cycle responses to elevated CO₂ and warming by regulating turnover rates of SOM pools and providing feedback pathways on C cycling, but very little work has been done in this area.

Mycorrhizal processes

Mycorrhizas will modify plant, community and ecosystem responses to global change factors. Functions mediated by mycorrhizas include plant nutrient foraging, plant C allocation and architecture, changes in soil structure, and soil C storage (Rillig et al., 2002; Staddon et al., 2002). Changes in atmospheric CO₂ concentrations indirectly affect mycorrhizas through changes in C allocation from host plants to fungi (Sanders et al., 1998; Treseder & Allen, 2000; Staddon et al., 2002; Olsrud et al., 2004), although mycorrhizal responses may be smaller when N availability is high (Treseder & Allen, 2000). Increased temperature may directly enhance arbuscular mycorrhizal (AM) colonization and development (Fitter *et al.*, 2000; Gavito et al., 2003). Indirect temperature responses may be mediated via changes in plant photosynthesis rate, plant and soil nutrient concentrations, and soil moisture. For example, ericoid mycorrhizal colonization increased with soil temperature due to an increased below-ground C allocation associated with low soil moisture content (Olsrud et al., 2004). In the same study, an interaction between elevated CO₂ and temperature increased plant water use efficiency, alleviating the soil moisture deficit and resulting in lower root and mycorrhizal densities compared to the effect of warming alone (Olsrud et al., 2004). Another study reported decreased AM mycorrhizal colonization under elevated temperature, and concluded that the negative effects of temperature on soil water and root production might have been offset by increased plant nutrient availability (Monz et al., 1994). Phosphorus uptake by AM mycorrhizas was stimulated by elevated temperatures more than by elevated CO₂, with no interaction, possibly because P uptake was not C limited (Gavito et al., 2003).

Soil and C pool sizes and turnover rates

Soil organic matter is comprised of a complex array of compounds with variable reactivity or susceptibility to decomposition. SOM has often been conceptually divided into two or three compartments which decompose rapidly, slowly, or not at all; these compartments have been called the active (labile, microbial), slow (intermediate, unprotected) and passive (recalcitrant, protected) pools, respectively (e.g. Schimel et al., 1985; Christensen, 1996, Fig. 2). Below-ground C pools can also include root biomass and litter or organic horizons lying above the mineral soil, each of which have different decomposition dynamics and responses to environmental change. Rapidly cycling, nonstructural carbohydrate pools, react quickly to disturbance or experimental manipulation, and can induce interannual variability in NEP (Hanson et al., 2003a). Long-term net changes in C storage resulting from elevated CO₂ or temperature require changes in slow-turnover pools, those with mean residence times of decades or longer. Therefore, knowing only the bulk C content of soil is insufficient; sizes and turnover rates of specific identifiable pools are required for predicting responses to environmental change.

Elevated CO₂ has stimulated NPP in most experiments to date, but the fate of this C, especially the portion allocated below-ground, largely remains unknown. Studies have rarely found measurable changes in SOM pools using conventional C analyses (e.g. Tate & Ross, 1997; Van Kessel et al., 2000; Leavitt et al., 2001). This has been attributed to the difficulty in measuring a small increment of SOC against a large background (Hungate et al., 1996). Stable C isotope labelling on both ambient and elevated CO₂ treatments during a FACE winter wheat experiment allowed detection of a net SOC increase of 5% over 2 years (Leavitt et al., 2001). In a semiarid grassland, new C inputs to the bulk soil were roughly doubled by twiceambient CO₂ over 4 years (Pendall et al., 2004b). However, increased turnover rates of older SOM negated the gain of new C, resulting in no difference in NEP between ambient and elevated treatments during moist conditions (Pendall et al., 2004a).

Warming often causes a rapid loss of labile substrates, which might make up c. 10% of the total SOM pool, followed by slower mineralization of intermediate SOM (Ineson et al., 1998a,b; Loiseau & Soussana, 1999; Melillo et al., 2002). A laboratory incubation experiment using Hawaiian soils and a ¹³C label (derived from C₃-C₄ vegetation shift) suggested that intermediate and active SOM pools had similar sensitivity to warming (Townsend et al., 1997).

Temperature-CO₂ experiments focusing on changes in below-ground C pool sizes and turnover rates have rarely been reported in the literature, reflecting the largest gap in our understanding of below-ground process responses to climate change. In a warming-CO₂-N experiment in tunnels with ryegrass swards, particulate organic matter increased under elevated CO₂, warming increased turnover rates, and the interaction of CO2 and warming strongly enhanced 'old' pool C decomposition (Loiseau & Soussana, 1999). In a shortgrass steppe study in which CO_2 concentrations were doubled, an interesting feedback was noted in that the Q_{10} for decomposition was lower under elevated CO_2 than ambient CO_2 (Pendall *et al.*, 2003). This reduced temperature sensitivity under elevated CO_2 suggests that substrate quality was diminished and/or that microbial community composition had shifted toward a greater importance of fungi, which have lower temperature response than bacteria.

Soil respiration and its components

Soil respiration – the diffusive flux of CO₂ (and CH₄) from the soil boundary layer into the atmosphere – is a functional and commonly used term. However, it is far too vague for use in below-ground process studies because there is no single process that defines what is measured (Fig. 2; Hanson et al., 2000). Under conditions that do not disturb the soil's natural surface boundary layer and microenvironment, measurements of CO₂ and CH₄ efflux from the soil surface are assumed to be in equilibrium with a wide range of below-ground biological processes. Those processes include autotrophic respiration associated with the growth and maintenance of roots and mycorrhizal fungi, rhizosphere microbial respiration tightly coupled to the supply of labile plant carbohydrates, and the respiration of heterotrophic decomposers. Because of the methodological difficulty in separating autotrophic root respiration from heterotrophic respiration by rhizosphere microbes, we define these combined processes as 'rhizosphere' respiration; 'decomposition', then, is the portion of the total soil CO₂ efflux that can be measured separately from the rhizosphere using mechanical or isotopic approaches (Hanson et al., 2000).

Autotrophic (rhizosphere) and heterotrophic respiratory processes are likely to respond to elevated temperature and CO₂ in different ways, and determination of the separate sources in multifactor studies is essential to a predictive understanding of ecosystem responses to global change. The majority of studies, however, have examined the effects of CO2 or warming on the total efflux rather than attempting to separate the components. Pajari (1995) studied *Pinus* spp. responses at 550 ppm CO_2 and a 2-3°C increase in temperature in opentop chambers, and found that increased soil respiration under elevated CO₂ was sometimes counteracted by increasing temperatures. Possibly, an indirect effect of warming led to drier soils, limiting respiration rates. Edwards & Norby (1999) evaluated the response of soil respiration under Acer to +300 ppm CO₂ and +4°C and found greater total soil respiration under each treatment, but the heterotrophic soil component was only increased by temperature and root growth/ activity was increased by both. Decomposition was more strongly stimulated by elevated CO₂ and warming together than by elevated CO₂ alone, but this interaction was dependent on adequate N supply in ryegrass swards (Loiseau & Soussana, 1999). Litter decomposition, rhizosphere respiration, and mineral soil respiration increased in experimental *Pseudotsuga* spp. mesocosms exposed to +200 ppm CO_2 and $+4^{\circ}C$ (Lin *et al.*, 1999). The decomposition component of soil respiration was only significantly affected by temperature, while rhizosphere respiration responded to both CO_2 and temperature.

Substrate availability, regulated by NPP, will ultimately limit the response of soil CO₂ efflux to altered conditions, regardless of the respiratory pathway. A key issue to consider is to what degree the increased supply of labile substrates under elevated CO2 enhances decomposition of pre-existing organic matter (i.e. 'priming'), and whether warming interacts with CO₂ to suppress or stimulate the priming effect. Over 2-3 yr of elevated CO₂, both wheat and shortgrass steppe showed increased decomposition under elevated CO₂, or priming (Pendall et al., 2001, 2003), but forest ecosystems with larger SOM pools may not show a priming effect. Norby et al. (2002) concluded that in forests, much of the C allocated below ground under elevated CO2 was entering a fast turnover pool, and insufficient experimental duration prevented build-up of mineral soil C to support a measurable increase in baseline heterotrophic decomposition rates. In wetlands, a portion of the plant carbon that enters the fast turnover pool is emitted as CH₄ (Whiting & Chanton, 2001).

Elevated CO_2 has been shown to increase CH_4 emissions from a variety of wetland ecosystems (Dacey *et al.*, 1994; Vann & Megonigal, 2003; and references therein), sometimes dramatically. This response effectively amplifies CO_2 radiative forcing by converting a portion of the CO_2 to CH_4 , a gas with a warming potential that is 8-21 times higher on a mole basis. Ecosystem respiration as CO_2 flux in peatlands appears more responsive to temperature than to changes in water table, while CH_4 fluxes in these systems are strongly responsive to water table fluctuations (Updegraff *et al.*, 2001). Interactions between elevated CO_2 and temperature on CH_4 emissions have not been investigated.

Decomposition of SOM is linked to N mineralization, providing feedbacks to NPP. Elevated CO₂ appears to have little effect on soil N mineralization (Norby *et al.*, 2001), but several studies have shown that soil warming can cause increased soil N mineralization and possibly nitrate leaching (Van Cleve *et al.*, 1990; Peterjohn *et al.*, 1994; Hobbie, 1996; Lükewille & Wright, 1997; Verburg *et al.*, 1999; Rustad *et al.*, 2001). In N-limited ecosystems, warming may relieve nutrient limitations on NPP under elevated CO₂ by increasing N mineralization (Shaver *et al.*, 2000). Litter decomposition is partly dependent on C: N ratios, which may be increased by elevated CO₂ and reduced by warming, but litter bag studies have so far shown little effect of these manipulations (Norby *et al.*, 2001).

Methodological Limitations and Suggestions for Improvements

Net ecosystem production

The long-term effects of elevated CO₂ and warming on C cycling may be predicted from changes in NEP, the difference

between gross primary production (GPP) and ecosystem respiration (R_e). If lateral transfers are ignored, NEP can be formulated as the difference between NPP and decomposition (R_h):

$$NEP = GPP - R_e = NPP - R_h$$
 Eqn 1

Often, decomposition is estimated from generic temperature response functions or mass loss over long periods and large regions. This approach severely restricts a mechanistic interpretation of responses to elevated CO_2 and warming, and more careful evaluation of drivers of decomposition rates is required to develop better predictive ability. A mass balance approach for estimating NEP in forest ecosystems that accounts for nonsteady state conditions is:

$$\begin{split} \text{NEP} &= (\text{NPP}_{\text{A}} - R_{\text{WD}}) + (\Delta C_{\text{FR}} + \\ &\quad \Delta C_{\text{CR}} + \Delta C_{\text{soil}} - \Delta C_{\text{litter}}) \end{split} \tag{Eqn 2}$$

Where NPP_A is above-ground net primary production, $R_{\rm WD}$ is the respiration from woody debris, $\Delta C_{\rm FR}$ is the net change in fine root C, $\Delta C_{\rm CR}$ is the difference between the net growth of live coarse roots and the decomposition of coarse roots attached to stumps, $\Delta C_{\rm soil}$ is the net change in mineral soil C, and $\Delta C_{\rm litter}$ is annual fine litter fall (Law *et al.*, 2003). The biometric approaches are extremely labour intensive, destructive, and still may miss a portion of C allocated below-ground. Where a stable isotope signal is present, NEP estimates may be constrained by using changes in $\delta^{13}{\rm C}$ of SOM to estimate rhizodeposition (Pendall *et al.*, 2004b) and $\delta^{13}{\rm C}$ of soil respiration to partition decomposition (Pendall *et al.*, 2003, 2004a).

Total below-ground C allocation

A simple budget approach can be used to estimate total below-ground C allocation (TBCA), the sum of C allocated below-ground for root and mycorrhizal respiration and turnover, and root exudates (Raich & Nadelhoffer, 1989, modified by Giardina & Ryan, 2002):

TBCA =
$$F_s - F_A + \Delta(C_{soil} + C_{litter} + C_{roots})$$
 Eqn 3

Where F_s is soil respiration and F_A is above-ground litter fall. Generally, these fluxes are evaluated over at least a year. Estimates of TBCA are useful for determining the total plant contribution to below-ground inputs (Giardina & Ryan, 2002; Giardina et al., 2003), for examining large-scale patterns in those inputs (Raich & Nadelhoffer, 1989; Davidson et al., 2002), and for constraining other estimates of below-ground activity based on measurements of the individual components (e.g. root turnover and respiration). Recently developed isotopic methods for evaluating root input and turnover rates (Gaudinski et al., 2001; Matamala et al., 2003) should be

applied cautiously because these estimates are sensitive to the presence of pre-treatment nonstructural carbohydrates (Luo, 2003).

Heterotrophic and autotrophic respiration

A key question is whether autotrophic and heterotrophic processes respond differently to warming and elevated CO₂; the heterotrophic flux is also required for estimates of NEP and thus C sequestration. Mechanical separation techniques of the respired sources have involved measurements of total soil CO₂ flux, then rhizosphere and litter respiration, with determination of heterotrophic respiration from soil by difference (Ryan et al., 1997; Law et al., 2001), girdling (Högberg et al., 2001), or root exclusion (Edwards & Norby, 1999). Stable isotope methods allow partitioning rhizosphere respiration and decomposition for elevated CO₂ experiments where isotopically distinct CO₂ is added (Pataki et al., 2003) and in areas that have transitioned between C₃ and C₄ cover (Rochette et al., 1999). This approach requires a minimum δ¹³C offset of about 4–5‰ between the currently growing biomass and pre-existing soil organic matter, which is not often found in C₃-dominated ecosystems unless a tracer has been added.

Below-ground C pools and turnover rates

The total amount of C in soil is very large in comparison to annual inputs or losses, and so for short duration experiments (< 5 years), changes in labile C pools are easier to detect than in longer-lived pools. Density separation results in a light fraction, which is recent, partially decomposed plant residue, and a heavy fraction, which is composed of older, organo-mineral complexes (Khanna *et al.*, 2002). Particle size separations have shown that organic matter in smaller size classes, associated with silt or clay, has lower turnover rates than larger, particulate organic matter (POM; Balesdent, 1996). Physical separation methods, however, often do not result in pools, that can be directly comparable to conceptual categories used in process models (W. Parton, personal communication).

Changes in soil C pool sizes and turnover rates in elevated CO₂ and warming experiments should be evaluated at a finer level of detail than has generally been done, possibly applying a combination of physical or chemical separation methods and stable isotope analyses, to better quantify small changes attributable to experimental conditions (e.g. Loiseau & Soussana, 1999). A limitation in most elevated CO₂ experiments is that a stable isotope tracer is present on the elevated, but not ambient, treatment, eliminating the possibility of comparing treatment effects without additional labelling (e.g. Leavitt *et al.*, 2001). Radiocarbon tracers provide important insights into climatically driven changes in turnover times, and can be applied to systems lacking a stable isotope label (Trumbore, 1997).

Data Synthesis using Models

In the past two decades, dozens of biogeochemical models have been developed to study ecosystem response to rising atmospheric CO₂ and global warming (e.g. Parton et al., 1987; Comins & McMurtrie, 1993; Rastetter et al., 1997; Luo & Reynolds, 1999; Thompson & Randerson, 1999; McGuire et al., 2001). Most of those models share a common structure that partitions photosynthetically fixed C into several pools. Although model complexity varies with numbers of pools and fluxes, modelling studies generally suggest that the predicted capacity of ecosystem C sequestration is strongly regulated by the residence time of C in these pools (Schimel et al., 1994; Joos et al., 1996; Luo & Reynolds, 1999; Thompson & Randerson, 1999). Thus, residence times of C in each of the pools, and their differential temperature sensitivities, are critical parameters for our predictive understanding of belowground responses to elevated CO₂ and warming. Verification of model results has been done by comparing SOC measurements over long-term (3-5 decades) field experiments with modelled output, showing generally good performance (Izaurralde et al., 2002). Although some below-ground models are constrained by available C stocks (e.g. Century (Parton et al., 1988) and Rothamsted (Jenkinson, 1990)) model performance of the mechanisms underlying C cycling dynamics at shorter time steps has rarely been evaluated.

Most terrestrial biogeochemistry models simulate exchanges between vegetation and soils that may influence the response of below-ground processes to changing CO₂ and temperature over time, and thus lead to CO₂ and temperature interactions (e.g. Cramer et al., 1999). These interactive effects are related to feedbacks mediated by another variable such as soil moisture, N availability, or litter quality. In most terrestrial biogeochemistry models, warming decreases soil moisture whereas elevated CO₂ increases it (e.g. Century, Pan et al., 1998, PnET, Aber et al., 1995). The changes in soil moisture have the potential to influence decomposition differentially along moisture gradients with larger relative effects in semiarid regions than in more mesic regions (Pan et al., 1998). The magnitude of these interaction effects depends on how hydrology is simulated and the assumed sensitivity of belowground processes to soil moisture. Interactions between CO₂ and nitrogen deposition have also been shown to influence modelled NEE, particularly when the effects of variable N availability during disturbance cycles are considered (Thornton et al., 2002).

Several terrestrial biogeochemistry models have incorporated the consequences of potential changes in litter quality on C sequestration into their simulations. The Century model allows C: N of leaf biomass to increase by 20% with a doubling of atmospheric CO₂, which decreased decomposition rates and N availability (Parton *et al.*, 1995). In a sensitivity study with the Terrestrial Ecosystem Model, McGuire *et al.* (1997) showed that the amount of soil organic C stored

across the globe was sensitive to potential changes in vegetation and litter C: N ratios associated with a doubling of atmospheric CO₂, as a result of both increased litter inputs and decreased sensitivity of decomposition to temperature caused by reduced litter quality. Field litter bag studies, however, rarely confirm model predictions of altered decomposition rates attributable to altered litter C: N ratios (Norby et al., 2001), suggesting another area for model validation.

Models should distinguish CO2 fluxes from roots versus heterotrophs, and respiration from surface organic and litter layers versus mineral soil horizons because the temporal dynamics of temperature, moisture, and nutrients vary spatially. C losses associated with the growth of new root tissues (root construction costs) should be modelled separately from seasonal temperature patterns as they can drive arbitrarily high estimates of temperature sensitivity (Boone et al., 1998; Hanson et al., 2003b). A few studies have attempted to reconcile modelled and measured values of decomposition and autotrophic respiration. Law et al. (2001) compared field estimates of annual heterotrophic respiration and autotrophic respiration from foliage, wood and roots (scaled up chamber measurements), ANPP, BNPP, and eddy flux measurements of total ecosystem respiration and NEE with Biome-BGC model outputs. The multiple measurements helped to identify areas for improvement in model assumptions, such as C allocation and fine root turnover rates. Pendall et al. (2003) compared decomposition simulated with an abiotic, empirical model with decomposition rates partitioned using stable isotopes, and found reasonably good agreement at ambient and elevated CO₂, allowing simulation of decomposition for additional growing seasons which lacked isotope data.

Synopsis

Recent improvements in below-ground techniques, such as estimation of root turnover rates, evaluation of 'new' C inputs and below-ground C allocation, partitioning soil respiration into root and microbial components, identification of C pools with distinct residence times, and increased attention to mycorrhizas, are bringing us closer to understanding belowground C cycling. Responses of soil C pools with turnover times of decades or longer will ultimately determine the net impact of climate change on below-ground C storage. The limited body of experimental evidence suggests that soil C cycling and decomposition may increase dramatically when warming and elevated CO₂ are combined. Long-term (decadal) responses will depend on whether substrate availability will be stimulated to the same degree, and whether substrate quality will be altered sufficiently to impact residence times of C pools. With few exceptions, however, current experiments do not adequately capture responses of slow pool C to altered environmental conditions.

Application of more standardized experimental methods in the field and laboratory would facilitate cross-site comparisons.

ID 1000 Death event a strong

Below-ground C dynamics could be evaluated more precisely using isotope pulse labelling or soil transplanting approaches in long-term (5–10 years), multifactor experiments. If experimental treatments are imposed on moisture gradients, responses mediated by soil water content may be detected. However, as the complexity and duration of experiments increase, the systems may become so perturbed that it is difficult to discriminate treatment effects from experimental artefacts. Intercomparable techniques for quantifying labile and slow C pool sizes have been in development, but quantifying residence times of these pools has been more difficult. Improved understanding of how the longer-lived C pools will react to anthropogenic climate change also requires validation of improved models.

Acknowledgements

The authors wish to thank Rich Norby and Lindsey Rustad for organizing the workshop 'Interactions between increasing CO₂ and temperature on terrestrial ecosystems', sponsored by GCTE, April, 2003. The lead author acknowledges substantial input from all coauthors who are listed alphabetically. Comments on an earlier draft from four anonymous reviewers improved the quality of the manuscript. Support for Paul Hanson's contribution to this paper was funded by the US Department of Energy's Office of Science (BER) through the Terrestrial Carbon Processes (TCP) program. The contributions of B. E. Law (Grant #FG0300ER63014) and J. P. Megonigal (Grant #DE-FG02–97ER62458) were supported by the Department of Energy.

References

- Aber JD, Ollinger SV, Federer CA, Reich PB, Goulden ML, Kicklighter DW, Melillo J, Lathrop RG Jr. 1995. Predicting the effects of climate change on water yield and forest production in the northeastern U.S. Climate Research 5: 207–222.
- Allen AS, Andrews JA, Finzi AC, Matamala R, Richter DD, Schlesinger WH. 2000. Effects of free-air CO_2 enrichment (FACE) on belowground processes in *Pinus taeda* forest. *Ecological Applications* 10: 437–448.
- Arnone JA III, Bohlen PJ. 1998. Stimulated $\rm N_2O$ flux from intact grassland monoliths afer two growing seasons under elevated atmospheric $\rm CO_2$. *Oecologia* 116: 331–335.
- Balesdent J. 1996. The significance of organic separates to carbon dynamics and its modelling in some cultivated soils. *European Journal of Soil Science* 47: 485–493.
- BassiriRad H. 2000. Kinetics of nutrient uptake by roots: responses to global change. *New Phytologist* 147: 155–169.
- BassiriRad H, Caldwell MM, Bilbrough C. 1993. Effects of soil temperature and nitrogen status on kinetics of ¹⁵NO₃⁻ uptake by roots of field-grown Agropyron desertorum (Fisch. Ex Link) Schult. New Phytologist 123: 485–489.
- Bassow SL, McConnaughay KDM, Bazzaz FA. 1994. The response of temperate tree seedlings grown in elevated CO₂ to extreme temperature events. *Ecological Applications* 4: 593–603.
- Berntson GM, Bazzaz FA. 1997. Nitrogen cycling in microcosms of yellow birch exposed to elevated CO₂: simultaneous positive and negative below-ground feedbacks. *Global Change Biology* 3: 247–258.

- Boone RD, Nadelhoffer KJ, Canary JD, Kaye JP. 1998. Roots exert a strong influence on the temperature sensitivity of soil respiration. *Nature* 396: 570–572.
- Christensen BT. 1996. Carbon in primary and secondary organomineral complexes. In: Carter MR, Stewart BA, eds. Structure and organic matter storage in agricultural soils. Boca Raton, FL, USA: Lewis Publishers, 97–165.
- Comins HN, McMurtrie RE. 1993. Long-term biotic response of nutrient-limited forest ecosystems to CO₂-enrichment: Equilibrium behavior of integrated plant-soil models. *Ecological Applications* 3: 666–681.
- Cotrufo MF, Ineson P, Scott A. 1998. Elevated CO₂ reduces the nitrogen concentration of plant tissues. Global Change Biology 4: 43–54.
- Cramer W, Kicklighter DW, Bondeau A, Moore B III, Churkina G, Nemry B, Ruimy A, Schloss AL. 1999. Comparing global models of terrestrial net primary productivity (NPP): Overview and key results. *Global Change Biology* 5: 1–15.
- Curtis PS. 1996. A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. *Plant, Cell & Evironment* 19: 127–137.
- Curtis PS, Balduman LM, Drake BG, Whigham DF. 1990. Elevated atmospheric CO₂ effects on belowground processes in C₃ and C₄ estuarine marsh communites. *Ecology* 71: 2001–2006.
- Curtis PS, Wang X. 1998. A meta-analysis of elevated CO₂ effects on woody plant mass, form and physiology. *Oecologia* 113: 299–313.
- Dacey JWH, Drake BG, Klug MJ. 1994. Stimulation of methane emission by carbon dioxide enrichment of marsh vegetation. *Nature* 370: 47–49.
- Davidson EA, Savage K, Bolstad P, Clark DA, Curtis PS, Ellsworth DS,
 Hanson PJ, Law BE, Luo Y, Pregitzer KS, Randolph JC, Zak D. 2002.
 Belowground carbon allocation in forests estimated from litterfall and
 IRGA-based soil respiration measurements. Agricultural and Forest
 Meteorology 113: 39–51.
- Diaz S, Grime JP, Harris J, McPherson E. 1993. Evidence of a feedback mechainsm limiting plant-response to elevated carbon-dioxide. *Nature* 364: 616–617.
- Edwards NT, Norby RJ. 1999. Below-ground respiratory responses of sugar maple and red maple saplings to atmospheric CO₂ enrichment and elevated air temperature. *Plant and Soil* 206: 85–97.
- Eissenstat DM, Wells CE, Yanai RD, Whitbeck JL. 2000. Building roots in changing environment: implications for root longevity. *New Phytologist* 147: 33–42.
- Fitter AH, Graves JD, Wolfenden J, Self GK, Brown TK, Bogie D, Mansfield TA. 1997. Root production and turnover and carbon budgets of two contrasting grasslands under ambient and elevated atmospheric carbon dioxide concentrations. New Phytologist 137: 247–255.
- Fitter AH, Graves JD, Wolfenden J, Self GK, Brown TK, Bogie D, Mansfield TA. 1999. Root production and turnover in an upland grassland subjected to artificial soil warming respond to radiation flux and nutrients, not temperature. *Oecologia* 120: 575–581.
- Fitter AH, Heinemeyer A, Staddon P. 2000. The impact of elevated CO₂ and global climate change on arbuscular mycorrhizas: a mycocentric approach. *New Phytologist* 147: 179–187.
- Forbes PJ, Black KE, Hooker JE. 1997. Temperature-induced alteration to root longevity in *Lolium perenne*. *Plant and Soil* 190: 87–90.
- Gaudinski JB, Trumbore SE, Davidson EA. 2001. The age of fine-root carbon in three forests of the eastern United States measured by radiocarbon. *Oecologia* 129: 420–429.
- Gavito ME, Schweiger P, Jakobsen I. 2003. P uptake by arbuscular mycorrhizal hyphae: effect of soil temperature and atmospheric CO₂ enrichment. Global Change Biology 9: 106–116.
- Giardina CP, Ryan MG. 2002. Total belowground carbon allocation in a fast-growing *Eucalyptus* plantation estimated using a carbon balance approach. *Ecosystems* 5: 487–499.
- Giardina CP, Ryan MG, Binkley D, Fownes JH. 2003. Primary production and carbon allocation in relation to nutrient supply in a tropical experimental forest. *Global Change Biology* 9: 1438–1450.

- Gill RA, Jackson RB. 2000. Global patterns of root turnover for terrestrial ecosystems. New Phytologist 147: 13-31.
- Hanson PJ, Edwards NT, Garten CT, Andrews JA. 2000. Separating root and soil microbial contributions to soil respiration: a review of methods and observations. Biogeochemistry 48: 115-146.
- Hanson PJ, Edwards NT, Tschaplinski TJ, Wullschleger SD, Joslin JD. 2003a. Estimating the net primary and net ecosystem production of a southeastern upland Quercus forest from an 8-year biometric record. In: Hanson, PJ, Wullschleger, SD, eds. North American Temperate Deciduous Forest Responses to Changing Precipitation Regimes. New York, USA: Springer, 378-395.
- Hanson PJ, O'Neill EG, Chambers ML, Riggs JS, Joslin JD, Wolfe MH. 2003b. Soil Respiration and Litter Decomposition. In: Hanson PJ, Wullschleger SD, eds. North American temperate deciduous forest responses to changing precipitation regimes. New York, USA: Springer, 163-189.
- Hendrick RL, Pregitzer KS. 1992. The demography of fine roots in a northern hardwook forest. Ecology 73: 1094-1104.
- Hendrick RL, Pregitzer KS. 1993. Patterns of fine-root mortality in two sugar maple forests. Nature 361: 59-61.
- Hendrick RL, Pregitzer KS. 1997. The relationship between fine-root demography and the soil environment in northern hardwood forests. Ecoscience 4: 99-105.
- Hobbie SE. 1996. Temperature and plant species control over litter decomposition in Alaskan tundra. Ecological Monographs 66:
- Högberg P, Nordgren A, Buchmann N, Taylor AFS, Ekblad A, Hogberg M, Nyberg G, Ottosson-Lofvenius M, Read DJ. 2001. Large-scale forest girdling shows that current photosynthesis drives soil respiration. Nature 411: 789-790.
- Hungate BA, Holland EA, Jackson RB, Chapin FSI, Mooney HA, Field CB. 1997. The fate of carbon in grasslands under carbon dioxide enrichment. Nature 388: 576-579.
- Hungate BA, Jackson RB, Field CB, Chapin FSI. 1996. Detecting changes in soil carbon in CO₂ enrichment experiments. Plant and Soil 187:
- Ineson P, Benham DG, Poskitt J, Harrison AF, Taylor K, Woods C. 1998b. Effects of climate change on nitrogen dynamics in upland soils. 2. A soil warming study. Global Change Biology 4: 153-161.
- Ineson P, Taylor K, Harrison AF, Poskitt J, Benham DG, Tipping E, Woof C. 1998a. Effects of climate change on nitrogen dynamics in upland soils. 1. A transplant approach. Global Change Biology 4: 143-152.
- IPCC. 2001. Summary for policymakers. In: Watson R, ed. Climate change 2001: the scientific basis. Cambridge, UK: Cambridge University Press.
- Izaurralde RC, Haugen-Kozyra KH, Jans DC, McGill WB, Grant RF, Hiley JC. 2002. Soil C dynamics: Measurement, simulation and site-to-region scale-up. In: Lal R, Kimble JM, Follett RF, Stewart BA, eds. Assessment methods for soil carbon. Boca Raton, FL, USA: Lewis Publishers, 553-575.
- Jackson RB, Mooney HA, Schulze ED. 1997. A global budget for fine-root biomass, surface area, and nutrient contents. Proceedings of the National Academy of Sciences, USA 94: 7362-7366.
- Jackson RB, Sala OE, Paruelo JM, Mooney HA. 1998. Ecosystem water fluxes for two grasslands in elevated ${\rm CO}_2$: a modeling analysis. Oecologia113: 537-546.
- Jenkinson DS. 1990. The turnover of organic carbon and nitrogen in soil. Philosophical Transactions of the Royal Society London Series B-Biology Sciences 329: 361-368.
- Jenkinson DS, Adams DE, Wild A. 1991. Model estimates of CO₂ emissions from soil in response to global warming. Nature 351:
- Jones TH, Thompson LJ, Lawton HJ, Bezemer TM, Bardgett RD, Blackburn TM, Bruce KD, Cannon PF, Hall GS, Hartley SE, Howson G, Hones CG, Mapichler C, Kandeler E, Ritchie DA. 1998. Impacts of rising atmospheric carbon dioxide on model terrestrial ecosystems. Science 280: 441-443.

- Joos F, Bruno M, Fink R, Siegenthaler U, Stocker TF, Le Quere C, Sarmiento JL. 1996. An efficient and accurate representation of complex oceanic and biospheric models of anthropogenic carbon uptake. Tellus 48B: 397-417
- Joslin JD, Wolfe MH. 1993. Temperature increase accelerates nitrate release from high-elevation red spruce soils. Canadian Journal of Forest Research
- Kandeler E, Tscherko D, Bardgett RD, Hobbs PJ, Lampichler C, Jones TH. 1998. The response of soil microorganisms and roots to elevated CO₂ and temperature in a terrestrial model ecosystem. Plant and Soil 2002: 251-262.
- Khanna PK, Ludwig B, Bauhus J, O'Hara C. 2002. Assessment and significance of labile organic C pools in forest soils. In: Lal R, Kimble JM, Follett RF, Stewart BA, eds. Assessment methods for soil carbon. Boca Raton, FL, USA: Lewis Publishers, 167-182.
- King JS, Pregitzer KS, Zak DR. 1999. Clonal variation in above- and belowground growth responses of Populus tremuloides Michaux: influence of soil warming and nutrient availability. Plant and Soil 217: 119-130.
- King JS, Thomas RB, Strain BR. 1996. Growth and carbon accumulation in root systems of Pinus taeda and Pinus ponderosa seedlings as affect by varying CO₂, temperature and nitrogen. *Tree Physiology* **16**: 635–642.
- King JS, Thomas RS, Strain BR. 1997. Morphology and tissue quality of seedling root systems of Pinus teada and Pinus ponderosa as affected by varying CO2, temperature and nitrogen. Plant and Soil 195:
- Kirschbaum MUF. 1995. The temperature dependence of soil organic matter decomposition, and the effect of global warming on soil organic C storage. Soil Biology and Biochemistry 27: 753-760.
- Kirschbaum MUF. 2000. Will changes in soil organic carbon act as a positive or negative feedback on global warming? Biogeochemistry 48:
- Law BE, Kelliher FM, Baldocchi DD, Anthoni PM, Irvine J. 2001. Spatial and temporal variation in respiration in a young ponderosa pine forest during a summer drought. Agricultural and Forest Meteorology 110: 27-43.
- Law BE, Sun OJ, Campbell JL, Van Tuyl S, Thornton PE. 2003. Changes in carbon storage and fluxes in a chronosequence of ponderosa pine. Global Change Biology 9: 510-524.
- Leavitt SW, Pendall E, Paul EA, Brooks T, Kimball BA, Pinter PJ, Johnson H, Wall G. 2001. Stable-carbon isotopes and soil organic carbon in the 1996 and 1997 FACE wheat experiments. New Phytologist 150: 305-314.
- Lenton TM, Huntingford C. 2003. Global terrestrial carbon storage and uncertainties in its temperature sensitivity examined with a simple model. Global Change Biology 9: 1333-1352.
- Lin G, Ehleringer JR, Lygiewicz PT, Johnson MG, Tingey DT. 1999. Elevated CO2 and temperature impacts on different components of soil CO₂ efflux in Douglas-fir terracosms. Global Change Biology 5: 157–168.
- Lloyd J, Taylor JA. 1994. On the temperature dependence of soil respiration. Functional Ecology 8: 315-323.
- Loiseau P, Soussana JF. 1999. Elevated [CO₂], temperature increase and N supply effects on the turnover of below-ground carbon in a temperate grassland ecosystem. Plant and Soil 210: 233-247.
- Long SP. 1991. Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO2 concentrations: has its importance been underestimated? Plant, Cell & Environment 14: 729-739.
- Lükewille A, Wright RF. 1997. Experimentally increased soil temperature causes release of nitrogen at a boreal forest catchment in southern Norway. Global Change Biology 3: 13-21.
- Luo Y. 2003. Uncertainties in interpretation of isotope signals for estimation of fine root longevity: theoretical considerations. Global Change Biology 9:
- Luo Y, Reynolds JF. 1999. Validity of extrapolating field CO₂ measurements to predict carbon sequestration in natural ecosystems. Ecology 80: 1568-1583.

- Matamala R, Gonzalez-Meler M, Jastrow J, Norby R, Schlesinger W. 2003. Impacts of fine root turnover on forest NPP and soil C sequestration potential. *Science* 302: 1385–1387.
- McGuire AD, Melillo J, Kicklighter DW, Pan Y, Xiao X, Helfrich J, Moore B, III Vorosmarty CJ, Schloss AL. 1997. Equilibrium responses of global net primary production and carbon storage to doubled atmospheric carbon dioxide: Sensitivity to changes in vegetation nitrogen concentration. Global Biogeochemical Cycles 11: 173–189.
- McGuire AD, Sitch S, Clein JS, Dargaville R, Esser G, Foley J, Heimann M, Joos F, Kaplan J, Kicklighter DW, Meier RA, Melillo JM, Moore B, III Prentice IC, Ramankutty N, Reichenau T, Schloss AL, Tian H, Williams LJ, Wittenberg U. 2001. Carbon balance of the terrestrial biosphere in the twentieth century: analyses of CO₂, climate and land use effects with four process-based ecosystem models. *Global Biogeochemical Cycles* 15: 183–206.
- McLain JET, Kepler TB, Ahmann DM. 2002. Belowground factors mediating changes in methane consumption in a forest soil under elevated CO₂. *Global Biogeochemical Cycles* **16**: 23–21–23–14.
- Megonigal JP, Schlesinger WH. 1997. Enhanced CH₄ emissions from a wetland soil exposed to elevated CO₂. *Biogeochemistry* 37: 77–88.
- Melillo JM, Steudler PA, Aber JD, Newkirk K, Lux H, Bowles FP, Catricala C, Magill A, Ahrens T, Morrisseau S. 2002. Soil warming and carbon-cycle feedbacks to the climate system. *Science* 298: 2173–2176.
- Monz CA, Kunt HW, Reeves FB, Elliot ET. 1994. The response of mycorrhizal colonization to elevated CO₂ and climate change in Pascopyrum smithii and Bouteloua gracilis. Plant and Soil 165: 75–80.
- Nelson JA, Morgan JA, LeCain DR, Mosier AR, Milchunas DG, Parton BA. 2004. Elevated CO₂ increases soil moisture and enhances plant water relations in a long-term field study in semi-arid shortgrass steppe of Colorado. *Plant and Soil*, in press.
- Norby RJ. 1994. Issues and perspectives for investigating root responses to elevated atmospheric carbon dioxide. *Plant and Soil* 165: 9–20.
- Norby RJ, Cotrufo MF, Ineson P, O'Neill EG, Canadell JG. 2001. Elevated CO₂, litter chemistry, and decomposition: a synthesis. *Oecologia* 127: 153–165.
- Norby RJ, Hanson PJ, O'Neill EG, Tschaplinski TJ, Weltzin JF, Hansen RA, Cheng W, Wullschleger SD, Gunderson CA, Edwards NT, Johnson DW. 2002. Net primary productivity of a CO₂-enriched deciduous forest and the implications for carbon storage. *Ecological Applications* 12: 1261–1266.
- Olsrud M, Melillo JM, Christensen TR, Michelsen A, Wallander H. 2004. Response of ericoid mycorrhizal colonisation and functioning to global change factors. *New Phytologist* 162: (In press.)
- Pajari B. 1995. Soil respiration in a poor upland site of Scots pine stand subjected to elevated temperatures and atmospheric carbon concentration. *Plant and Soil* 168–169: 563–570.
- Pan Y, Melillo J, McGuire AD, Kicklighter DW, Pitelka LF, Hibbard K, Pierce LL, Running SW, Ojima D, Parton WJ, Other members of the Vemap working Group. 1998. Modeled responses of terrestrial ecosystems to elevated atmospheric CO₂: a comparison of simulations by the biogeochemistry models of the Vegetation/Ecosystem Modeling and Analysis Project (VEMAP). *Oecologia* 114: 389–404.
- Parton WJ, Schimel DS, Cole CV, Ojima DS. 1987. Analysis of factors controlling soil organic matter levels in Great Plains grasslands. Soil Science Society of America Journal 51: 1173–1179.
- Parton WJ, Scurlock JM, Ojima DS, Schimel DS, Hall DO, Members of the ScopeGram Group. 1995. Impact of climate change on grassland production and soil carbon worldwide. Global Change Biology 1: 13–22.
- Parton WJ, Stewart JWB, Cole CV. 1988. Dynamics of C, N, P and S in grassland soils: a model. *Biogeochemistry* 5: 109–131.
- Pataki DE, Ellsworth DS, Evans LD, Gonzalez-Meler M, King J, Leavitt SW, Lin G, Matamala R, Pendall E, Siegwolf R, Van Kessel C, Ehleringer JR. 2003. Tracing changes in ecosystem function under elevated carbon dioxide conditions. *Bioscience* 53: 805–818.

- Pendall E, Del Grosso S, King JY, LeCain DR, Milchunas DG, Morgan JA, Mosier AR, Ojima D, Parton WA, Tans PP, White JWC. 2003. Elevated atmospheric CO₂ effects and soil water feedbacks on soil respiration components in a Colorado grassland. *Global Biogeochemical Cycles* 17: doi: 10.1029/2001GB001821.
- Pendall E, King JY, Mosier AR, Morgan JA, Milchunas DS. 2004a.

 Stable isotope constraints on net ecosystem production in elevated CO₂ experiments. In: Flanagan LB, Ehleringer JR, Pataki DE, eds. *Stable isotopes and biosphere-atmosphere interactions* (In press.)
- Pendall E, Leavitt SW, Brooks T, Kimball BA, Pinter PJ Jr, Wall GW, LaMorte R, Wechsung G, Wechsung F, Adamsen F, Matthias AD, Thompson TL. 2001. Elevated CO₂ stimulates soil respiration in a FACE wheat field. *Basic and Applied Ecology* 2: 193–201.
- Pendall E, Mosier AR, Morgan JA. 2004b. Rhizodeposition stimulated by elevated CO₂ in a semi-arid grassland. *New Phytologist* 162: (In press.)
- Peterjohn WT, Melillo J, Streudler PM, Newkirk KM, Bowles FP, Aber JD. 1994. Responses of trace gas fluxes and N availability to experimentally elevated soil temperatures. *Ecological Applications* 4: 617–625
- Pregitzer KS, DeForest JL, Burton AJ, Allen MF, Ruess RW, Hendrick RL. 2002. Fine-root architecture of nine North American trees. *Ecological Monographs* 72: 293–309.
- Pregitzer KS, Laskowski MJ, Burton AJ, Lessard VC, Zak DR. 1998.
 Variation in northern hardwood root respiration with root diameter and soil depth. *Tree Physiology* 18: 665–670.
- Pregitzer KS, Zak DR, Maziasz J, DeForest J, Curtis PS, Lussenhop J. 2000. Fine-root growth, mortality, and morphology in a factorial elevated atmospheric CO₂ soil N availability experiment. *Ecological Applications* 10: 18–33.
- Raich JW, Nadelhoffer KJ. 1989. Belowground carbon allocation in forest ecosystems: global trends. *Ecology* 70: 1346–1354.
- Raich JW, Potter CS, Bhagawati D. 2002. Interannual variability in global soil respiration, 1980–94. Global Change Biology 8: 800–812.
- Raich JW, Schlesinger WH. 1992. The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus* 44B: 81–99.
- Rastetter EB, Agren GI, Shaver GR. 1997. Responses of N-limited ecosystems to increased CO₂: a balanced-nutrition, coupled-element-cycles model. *Ecological Applications* 7: 444–460.
- Rillig MC, Treseder KK, Allen MF. 2002. Global change and mycorrhizal fungi. In: Heijden MGA, Sanders IR, eds. *Mycorrhizal ecology*. New York, USA: Springer-Verlag, 135–160.
- Rochette P, Flanagan LB, Gregorich EG. 1999. Seperating Soil Respiration into Plant and Soil Components Using Analysis of the Natural Abundance of Carbon-13. Soil Science Society of America Journal 63: 1207–1213.
- Rogers HH, Runion GB, Prior SA, Torbert HA. 1999. Responses of plants to elevated atmospheric CO₂: root growth, mineral nutrition and soil carbon. In: Luo Y, Mooney HA, eds. *Carbon dioxide and environmental stress*. San Diego, CA, USA: Academic Press, 215–244.
- Ronn R, Gavito M, Larsen J, Jakobsen I, Frederiksen H, Christensen S. 2002. Response of free-living soil protozoa and microorganisms to elevated atmospheric CO₂ and presence of mycorrhizae. Soil Biology and Biochemistry 34: 923–932.
- Rustad LE, Campbell JL, Marion GM, Norby RJ, Mitchell MJ, Hartley AE, Cornelissen JHC, Gurevitch J. 2001. A meta-analysis of the response of soil respiration, net nitrogen mineralization, and aboveground plant growth to experimental ecosystem warming. *Oecologia* 126: 542–562.
- Ryan MG, Hubbard RM, Pongracic S, Raison RJ, McMurtrie RE. 1996. Foliage, fine-root, woody-tissue and stand respiration in *Pinus radiata* in relation to nutrient status. *Tree Physiology* 16: 333–343.
- Ryan MG, Lavigne MB, Gower ST. 1997. Annual carbon cost of autotrophic respiration in boreal forest ecosystems in relation to species and climate. *Journal of Geophysical Research* 102: 28871–28883.

- Saleska SR, Harte J, Torn MS. 1999. The effect of experimental ecosystem warming on CO₂ fluxes in a montane meadow. Global Change Biology 5: 125 - 141.
- Sanders IR, Streitwolf-Engel R, van der Heijden MGA, Boller T, Wiemken A. 1998. Increased allocation to external hyphae of arbuscular mycorrhizal fungi under CO2 enrichment. Oecologia 117: 496-503.
- Schimel D, Braswell BH, Holland EA, McKeown R, Ojima D, Painter TH, Parton WJ, Townsend AR. 1994. Climatic, edaphic, and biotic controls over storage and turnover of carbon in soils. Global Biogeochemical Cycles 8. 279-293
- Schimel D, Coleman DC, Horton KA. 1985. Soil organic matter dynamics in paired rangeland and cropland toposequences in North Dakota. Geoderma 36: 201-214.
- Shaver GR, Canadell FS, Chapin FS, IIIGurevitch J, Harte J, Henry G, Ineson P, Jonasson S, Melillo J, Pitelka L, Rustad L. 2000. Global warming and terrestrial ecosystems: a conceptual framework for analysis. Bioscience 50: 871-882.
- Smart DR, Ritchie K, Stark JM, Bugbee B. 1997. Evidence that elevated CO, levels can indirectly increase rhizosphere denitrifier activity. Applied and Environmental Microbiology 63: 4621-4624.
- Soussana J, Casella E, Loiseau P. 1996. Long-term effects of CO₂ enrichment and temperature increase on a temperate grass sward. II. Plant nitrogen budgets and root fraction. Plant and Soil 182: 101-114.
- Staddon PL, Heinemeyer A, Fitter AH. 2002. Mycorrhizas and global environmental change: research at different scales. Plant and Soil 244:
- Strömgren M, Linder S. 2002. Effects of nutrition and soil warming on stemwood production in a boreal Norway spruce stand. Global Change Biology 8: 1195-1204.
- Tate KR, Ross DJ. 1997. Elevated CO2 and moisture effects on soil carbon storage and cycling in temperate grasslands. Global Change Biology
- Thompson MV, Randerson JT. 1999. Impulse response functions of terrestrial carbon cycle models: Method and application. Global Change Biology 5: 371-394.
- Thornton PE, Law BE, Gholz HL, Clark KL, Falge E, Ellsworth DS, Goldstein AH, Monson RK, Hollinger D, Falk M, Chen J, Sparks JP. 2002. Modeling and measuring the effects of disturbance history and climate on carbon and water budgets in evergreen needleleaf forests. Agricultural and Forest Meteorology 113: 185-222.

- Townsend AR, Vitousek PM, Desmarais DJ, Tharpe A. 1997. Soil carbon pool structure and temperature sensitivity inferred using CO₂ and ¹³CO₂ incubation fluxes from five Hawaiian soils. Biogeochemistry 38: 1-17.
- Treseder KK, Allen MF. 2000. Mycorrhizal fungi have a potential role in soil carbon storage under elevated CO2 and nitrogen deposition. New Phytologist 147: 189-200.
- Trumbore SE. 1997. Potential responses of soil organic carbon to global environmental change. Proceedings of the National Academy of Sciences, USA 94: 8284-8291.
- Updegraff K, Bridgham SD, Pastor J, Weishampel PHC. 2001. Ecosystem respiration response to warming and water-table manipulations in peatland mesocosms. Ecological Applications 11: 311–326.
- Van Cleve K, Oechel WD, Hom JL. 1990. Response of black spruce (Picea mariana) ecosystems to soil temperature modification in interior Alaska. Canadian Journal of Forest Research 20: 1530-1535.
- Van Kessel C, Nitschelm J, Horwath WR, Harris D, Walley F, Luscher A, Hartwig U. 2000. Carbon-13 input and turn-over in a pasture soil exposed to long-term elevated atmospheric CO2. Global Change Biology 6:
- Vann C, Megonigal J. 2003. Elevated CO₂ and water depth regulation of methane emissions: Comparison of woody and non-woody wetland plant species. Biogeochemistry 63: 117-134.
- Verburg PSJ, Van Loon WKP, Lükewille A. 1999. The CLIMEX experiment: soil response after 2 years of treatment. Biology and Fertility of Soils 28: 271-276.
- Wan S, Norby RJ, Pregitzer KS, Ledford J, O'Neill EG. 2004. CO₂ enrichment and warming of the atmosphere enhance both productivity and mortality of maple tree fine roots. New Phytologist 162: (In press.)
- Whiting GJ, Chanton JP. 2001. Greenhouse carbon balance of wetlands: methane emission versus carbon sequestration. Tellus 53B:
- Zak DR, Holmes WE, Curtis PS, Teeri JA, Fogel R, Randlett DL. 1993. Elevated atmospheric CO2 and feedback between and nitrogen cycles. Plant and Soil 151: 105-117.
- Zak DR, Pregitzer KS, Curtis PS, Teeri JA, Fogel R, Randlett DL. 2000a. Atmospheric CO2 and the composition and function of soil microbial communities. Ecological Applications 10: 47-59.
- Zak DR, Pregitzer KS, King JS, Holmes WE. 2000b. Elevated atmospheric CO2, fine roots and the response of soil microorganisms: a review and hypothesis. New Phytologist 147: 201-222.



About New Phytologist

- New Phytologist is owned by a non-profit-making charitable trust dedicated to the promotion of plant science, facilitating projects from symposia to open access for our Tansley reviews. Complete information is available at www.newphytologist.org
- Regular papers, Letters, Research reviews, Rapid reports and Methods papers are encouraged. We are committed to rapid processing, from online submission through to publication 'as-ready' via OnlineEarly - average first decisions are just 5-6 weeks. Essential colour costs are free, and we provide 25 offprints as well as a PDF (i.e. an electronic version) for each article.
- For online summaries and ToC alerts, go to the website and click on 'Journal online'. You can take out a personal subscription to the journal for a fraction of the institutional price. Rates start at £108 in Europe/\$193 in the USA & Canada for the online edition (click on 'Subscribe' at the website)
- If you have any questions, do get in touch with Central Office (newphytol@lancaster.ac.uk; tel +44 1524 592918) or, for a local contact in North America, the USA Office (newphytol@ornl.gov; tel 865 576 5261)